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PII: S0092-8674(21)00991-0

DOI: <https://doi.org/10.1016/j.cell.2021.08.017>

Reference: CELL 12163

To appear in: *Cell*

Please cite this article as: Holmes, E.C., Goldstein, S.A., Rasmussen, A.L., Robertson, D.L., Crits-Christoph, A., Wertheim, J.O., Anthony, S.J., Barclay, W.S., Boni, M.F., Doherty, P.C., Farrar, J., Geoghegan, J.L., Jiang, X., Leibowitz, J.L., Neil, S.J.D., Skern, T., Weiss, S.R., Worobey, M., Andersen, K.G., Garry, R.F., Rambaut, A., The Origins of SARS-CoV-2: A Critical Review, *Cell* (2021), doi: <https://doi.org/10.1016/j.cell.2021.08.017>.

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The Origins of SARS-CoV-2: A Critical Review

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Since the first reports of a novel SARS-like coronavirus in December 2019 in Wuhan, China, there has been intense interest in understanding how SARS-CoV-2 emerged in the human population. Recent debate has coalesced around two competing ideas: a “laboratory escape” scenario and zoonotic emergence. Here, we critically review the current scientific evidence that may help clarify the origin of SARS-CoV-2.

Evidence supporting a zoonotic origin of SARS-CoV-2

Coronaviruses have long been known to present a high pandemic risk. SARS-CoV-2 is the ninth documented coronavirus that infects humans and the seventh identified in the last 20 years (Lednický et al., 2021; Vlasova et al., 2021). All previous human coronaviruses have zoonotic origins, as have the vast majority of human viruses. The emergence of SARS-CoV-2 bears several signatures of these prior zoonotic events. It displays clear similarities to SARS-CoV that spilled over into humans in Foshan, Guangdong province, China in November 2002, and again in Guangzhou, Guangdong province in 2003 (Xu et al., 2004). Both these SARS-CoV emergence events were associated with markets selling live animals and involved species, particularly civets and raccoon dogs (Guan et al., 2003), that were also sold live in Wuhan markets in 2019 (Xiao et al., 2021) and are known to be susceptible to SARS-CoV-2 infection (Freuling et al., 2020). Animal traders working in 2003, without a SARS diagnosis, were documented to have high levels of IgG to SARS-CoV (13% overall and >50% for traders specializing in civets; Centers for Disease Control and Prevention, 2003). Subsequent serological surveys found ~3% positivity rates to SARS-related coronaviruses (SARSr-CoV) in residents of Yunnan province living close to bat caves (Wang et al., 2018), demonstrating regular exposure in rural locations. The closest known relatives to both SARS-CoV and SARS-CoV-2 are viruses from bats in Yunnan, although animals from this province have been preferentially sampled. For both SARS-CoV and SARS-CoV-2 there is a considerable geographic gap between Yunnan and the location of the first human cases, highlighting the difficulty in identifying the exact pathway of virus emergence and the importance of sampling beyond Yunnan.

SARS-CoV-2 also shows similarities to the four endemic human coronaviruses: HCoV-OC43, HCoV-HKU1, HCoV-229E, and HCoV-NL63. These viruses have zoonotic origins and the

circumstances of their emergence are unclear. In direct parallel to SARS-CoV-2, HCoV-HKU1, which was first described in a large Chinese city (Shenzhen, Guangdong) in the winter of 2004, has an unknown animal origin, contains a furin cleavage site in its spike protein, and was originally identified in a case of human pneumonia (Woo et al., 2005).

Based on epidemiological data, the Huanan market in Wuhan was an early and major epicenter of SARS-CoV-2 infection. Two of the three earliest documented COVID-19 cases were directly linked to this market selling wild animals, as were 28% of all cases reported in December 2019 (WHO, 2021). Overall, 55% of cases during December 2019 had an exposure to either the Huanan or other markets in Wuhan, with these cases more prevalent in the first half of that month (WHO, 2021). Examination of the locations of early cases shows that most cluster around the Huanan market, located north of the Yangtze river (**Figure 1B-E**), although case reporting may be subject to sampling biases reflecting the density and age structure of the population in central Wuhan, and exact location of some early cases is uncertain. These districts were also the first to exhibit excess pneumonia deaths in January 2020 (**Figure 1F-H**), a metric that is less susceptible to the potential biases associated with case reporting. There is no epidemiological link to any other locality in Wuhan, including the Wuhan Institute of Virology (WIV) located south of the Yangtze and the subject of considerable speculation. Although some early cases do not have a direct epidemiological link to a market (WHO, 2021), this is expected given high rates of asymptomatic transmission and undocumented secondary transmission events, and was similarly observed in early SARS-CoV cases in Foshan (Xu et al., 2004).

During 2019, markets in Wuhan – including the Huanan market – traded many thousands of live wild animals including high-risk species such as civets and raccoon dogs (Xiao et al., 2021). Following its closure, SARS-CoV-2 was detected in environmental samples at the Huanan market, primarily in the western section that traded in wildlife and domestic animal products, as well as in associated drainage areas (WHO, 2021). While animal carcasses retrospectively tested negative for SARS-CoV-2, these were unrepresentative of the live animal species sold, and specifically did not include raccoon dogs and other animals known to be susceptible to SARS-CoV-2 (Xiao et al., 2021).

The earliest split in the SARS-CoV-2 phylogeny defines two lineages - denoted A and B (Rambaut et al., 2020) - that likely circulated contemporaneously (**Figure 1A**). Lineage B, which became dominant globally, was observed in early cases linked to the Huanan market and environmental samples taken there, while lineage A contains a case with exposure to other markets (**Figure 1A-B**) as well as with later cases in Wuhan and other parts of China (WHO, 2021). This phylogenetic pattern is consistent with the emergence of SARS-CoV-2 involving one or more contacts with infected animals and/or traders, including multiple spill-over events, as potentially infected or susceptible animals were moved into or between Wuhan markets via shared supply chains and sold for human consumption (Xiao et al., 2021). The potential emergence of SARS-CoV-2 across multiple markets again mirrors SARS-CoV in which high levels of infection, seroprevalence and genetic diversity in animals were documented at both the Dongmen market in Shenzhen (Al, 2004; Guan et al., 2003) and the Xinyuan market in Guangzhou (Tu et al., 2004; Wang et al., 2005).

Viruses closely related to SARS-CoV-2 have been documented in bats and pangolins in multiple localities in South-East Asia, including in China, Thailand, Cambodia, and Japan (Lytras et al. 2021; Zhou et al., 2021), with serological evidence for viral infection in pangolins for more than a decade (Wacharapluesadee et al., 2021). However, a significant evolutionary gap exists between SARS-CoV-2 and the closest related animal viruses: for example, the bat virus RaTG13 collected by the WIV has a genetic distance of approximately 4% (~1,150 mutations) to the Wuhan-Hu-1 reference sequence of SARS-CoV-2, reflecting decades of evolutionary divergence (Boni et al., 2020). Widespread genomic recombination also complicates the assignment of which viruses are closest to SARS-CoV-2. Although RaTG13, sampled from a *Rhinolophus affinis* bat in Yunnan (Zhou et al., 2020b), has the highest average genetic similarity to SARS-CoV-2, a history of recombination means that three other bat viruses – RmYN02, RpYN06 and PrC31 – are closer in most of the virus genome (particularly ORF1ab) and thus share a more recent common ancestor with SARS-CoV-2 (Li et al., 2021; Lytras et al. 2021; Zhou et al., 2021). None of these three closer viruses were collected by the WIV and all were sequenced after the pandemic had begun (Li et al., 2021; Zhou et al., 2020a; Zhou et al., 2021). Collectively, these data demonstrate beyond reasonable doubt that RaTG13 is not the progenitor of SARS-CoV-2, with or without laboratory manipulation or experimental mutagenesis.

No bat reservoir nor intermediate animal host for SARS-CoV-2 has been identified to date. This is presumably because the right animal species and/or populations have not yet been sampled and/or any progenitor virus may be at low prevalence. Initial cross-species transmission events are also very likely to go undetected. Most SARS-CoV-2 index case infections will not have resulted in sustained onward transmission (Pekar et al., 2021) and only a very small fraction of spillovers from animals to humans result in major outbreaks. Indeed, the animal origins of many well-known human pathogens, including Ebola virus, Hepatitis C virus, poliovirus, and the coronaviruses HCoV-HKU1 and HCoV-NL63, are yet to be identified, while it took over a decade to discover bat viruses with >95% similarity to SARS-CoV and able to use hACE-2 as a receptor (Hu et al., 2017).

Could SARS-CoV-2 have escaped from a laboratory?

There are precedents for laboratory incidents leading to isolated infections and transient transmission chains, including SARS-CoV (Parry, 2004). However, with the exception of Marburg virus (Ristanović et al., 2020), all documented laboratory escapes have been of readily identifiable viruses capable of human infection and associated with sustained work in high titer cultures (Geddes, 2006; Lim et al., 2004; Senio, 2003). The 1977 A/H1N1 influenza pandemic, that most likely originated from a large-scale vaccine challenge trial (Rozo and Gronvall, 2015), is the only documented example of a human epidemic or pandemic resulting from research activity. No epidemic has been caused by the escape of a novel virus and there is no data to suggest that the WIV—or any other laboratory—was working on SARS-CoV-2, or any virus close enough to be the progenitor, prior to the COVID-19 pandemic. Viral genomic sequencing without cell culture, which was routinely performed at the WIV, represents a negligible risk as viruses are inactivated during RNA extraction (Blow et al., 2004). No case of laboratory escape has been documented following the sequencing of viral samples.

Known laboratory outbreaks have been traced to both workplace and family contacts of index cases and to the laboratory of origin (Geddes, 2006; Lim et al., 2004; Ristanović et al., 2020; Senio, 2003). Despite extensive contact tracing of early cases during the COVID-19 pandemic, there have been no reported cases related to any laboratory staff at the WIV and all staff in the

laboratory of Dr. Shi Zhengli were said to be seronegative for SARS-CoV-2 when tested in March 2020 (WHO, 2021), with the laboratory reportedly following the appropriate biosafety protocols during their coronavirus work (Cohen, 2020). During a period of high influenza transmission and other respiratory virus circulation (Liu et al., 2020a) reports of illnesses would need to be confirmed as caused by SARS-CoV-2 to be relevant. Epidemiological modeling suggests that the number of hypothetical cases needed to result in multiple hospitalized COVID-19 patients prior to December 2019 is incompatible with observed clinical, genomic, and epidemiological data (Pekar et al., 2021).

The WIV possesses an extensive catalogue of samples derived from bats (Latinne et al., 2020) and has reportedly successfully cultured three SARSr-CoVs from bats – WIV1, WIV16 and Rs4874 (Ge et al., 2013; Hu et al., 2017; Yang et al., 2015). Importantly, all three viruses are more closely related to SARS-CoV than to SARS-CoV-2 (Ge et al., 2013; Hu et al., 2017; Yang et al., 2015). In contrast, bat virus RaTG13 from the WIV has reportedly never been isolated nor cultured and only exists as a nucleotide sequence assembled from short sequencing reads (Cohen, 2020). The three cultured viruses were isolated from fecal samples through serial amplification in Vero E6 cells, a process that consistently results in the loss of the SARS-CoV-2 furin cleavage site (Davidson et al., 2020; Klimstra et al., 2020; Liu et al., 2020b; Ogando et al., 2020; Sasaki et al., 2021; Wong et al., 2020; Zhu et al., 2021b). It is therefore highly unlikely that these techniques would result in the isolation of a SARS-CoV-2 progenitor with an intact furin cleavage site. No published work indicates that other methods, including the generation of novel reverse genetics systems, were used at the WIV to propagate infectious SARSr-CoVs based on sequence data from bats. Gain-of-function research would be expected to utilize an established SARSr-CoV genomic backbone, or at a minimum a virus previously identified via sequencing. However, past experimental research using recombinant coronaviruses at the WIV has used a genetic backbone (WIV1) unrelated to SARS-CoV-2 (Hu et al., 2017) and SARS-CoV-2 carries no evidence of genetic markers one might expect from laboratory experiments (Andersen et al., 2020). There is no rational experimental reason why a new genetic system would be developed using an unknown and unpublished virus, with no evidence nor mention of a SARS-CoV-2-like virus in any prior publication or study from the WIV (Ge et al., 2012; Hu et al., 2017; Menachery et al., 2015), no evidence that the WIV sequenced a virus that is closer to

SARS-CoV-2 than RaTG13, and no reason to hide research on a SARS-CoV-2-like virus prior to the COVID-19 pandemic. Under any laboratory escape scenario SARS-CoV-2 would have to have been present in a laboratory prior to the pandemic, yet no evidence exists to support such a notion and no sequence has been identified that could have served as a precursor.

A specific laboratory escape scenario involves accidental infection in the course of serial passage of a SARS-CoV in common laboratory animals such as mice. However, early SARS-CoV-2 isolates were unable to infect wild-type mice (Wan et al., 2020). While murine models are useful for studying infection *in vivo* and testing vaccines, they often result in mild or atypical disease in hACE2 transgenic mice (Bao et al., 2020; Hassan et al., 2020; Israelow et al., 2020; Rathnasinghe et al., 2020; Sun et al., 2020b). These findings are inconsistent with a virus selected for increased pathogenicity and transmissibility through serial passage through susceptible rodents. Although SARS-CoV-2 has since been engineered (Dinnon et al., 2020) and mouse-adapted by serial passage (Gu et al., 2020; Leist et al., 2020; Sun et al., 2020a), specific mutations in the spike protein, including N501Y, are necessary for such adaptation in mice (Gu et al., 2020; Sun et al., 2020a). Notably, N501Y has arisen convergently in multiple SARS-CoV-2 variants of concern in the human population, presumably being selected to increase ACE2 binding affinity (Khan et al., 2021; Kuzmina et al., 2021; Liu et al., 2021; Starr et al., 2020). If SARS-CoV-2 resulted from attempts to adapt a SARS-CoV for study in animal models, it would likely have acquired mutations like N501Y for efficient replication in that model, yet there is no evidence to suggest such mutations existed early in the pandemic. Both the low pathogenicity in commonly used laboratory animals and the absence of genomic markers associated with rodent adaptation indicate that SARS-CoV-2 is highly unlikely to have been acquired by laboratory workers in the course of viral pathogenesis or gain-of-function experiments.

Evidence from genomic structure and ongoing evolution of SARS-CoV-2

Considerable attention has been devoted to claims that SARS-CoV-2 was genetically engineered or adapted in cell culture or “humanized” animal models to promote human transmission (Zhan et al., 2020). Yet, since its emergence, SARS-CoV-2 has experienced repeated sweeps of mutations that have increased viral fitness (Deng et al., 2021; Otto et al., 2021; Simmonds,

2020). The first clear adaptive mutation, the D614G substitution in the spike protein, occurred early in the pandemic (Korber et al., 2020; Volz et al., 2021). Recurring mutations in the receptor binding domain of the spike protein, including N501Y, K417N/T, L452R, and E484K/Q—constituent mutations of the variants of concern—similarly enhance viral infectivity (Cai et al., 2021; Khan et al., 2021; Kuzmina et al., 2021) and ACE2 binding (Liu et al., 2021; Starr et al., 2020; Zhu et al., 2021a), refuting claims that the SARS-CoV-2 spike protein was optimized for binding to human ACE2 upon its emergence (Piplani et al., 2021). Further, some pangolin-derived coronaviruses have receptor binding domains that are near-identical to SARS-CoV-2 at the amino acid level (Andersen et al., 2020; Xiao et al., 2020) and bind to human ACE2 even more strongly than SARS-CoV-2, showing that there is capacity for further human adaptation (Dicken et al., 2021). SARS-CoV-2 is also notable for being a host generalist virus (Conceicao et al., 2020), capable of efficient transmission in multiple mammalian species, including mink, tigers, cats, gorillas, dogs, raccoon dogs, ferrets, and large outbreaks have been documented in mink with spill-back to humans (Oude Munnink et al., 2021) and to other animals (van Aart et al., 2021). Combined, these findings show that no specific human “pre” adaptation was required for the emergence or early spread of SARS-CoV-2, and the claim that the virus was already highly adapted to the human host (Zhan et al., 2020), or somehow optimized for binding to human ACE2, is without validity.

The genesis of the polybasic (furin) cleavage site in the spike protein of SARS-CoV-2 has been subject to recurrent speculation. Although the furin cleavage site is absent from the closest known relatives of SARS-CoV-2 (Andersen et al., 2020), this is unsurprising as the lineage leading to this virus is poorly sampled and the closest bat viruses have divergent spike proteins due to recombination (Boni et al., 2020; Lytras et al. 2020; Zhou et al., 2021). Furin cleavage sites are commonplace in other coronavirus spike proteins, including some feline alphacoronaviruses, MERS-CoV, most but not all strains of mouse hepatitis virus, as well as in endemic human betacoronaviruses such as HCoV-OC43 and HCoV-HKU1 (Gombold et al., 1993; de Haan et al., 2008; Kirchdoerfer et al., 2016). A near identical nucleotide sequence is found in the spike gene of the bat coronavirus HKU9-1 (Gallaher, 2020), and both SARS-CoV-2 and HKU9-1 contain short palindromic sequences immediately upstream of this sequence that are indicative of natural recombination break-points via template switching (Gallaher, 2020).

Hence, simple evolutionary mechanisms can readily explain the evolution of an out-of-frame insertion of a furin cleavage site in SARS-CoV-2 (**Figure 2**).

The SARS-CoV-2 furin cleavage site (containing the amino acid motif RRAR) does not match its canonical form (R-X-R/K-R), is suboptimal compared to those of HCoV-HKU1 and HCoV-OC43, lacks either a P1 or P2 arginine (depending on the alignment), and was caused by an out-of-frame insertion (**Figure 2**). The RRAR and RRSR S1/S2 cleavage sites in feline coronaviruses (FCoV) and cell-culture adapted HCoV-OC43, respectively, are not cleaved by furin (de Haan et al., 2008). There is no logical reason why an engineered virus would utilize such a suboptimal furin cleavage site, which would entail such an unusual and needlessly complex feat of genetic engineering. The only previous studies of artificial insertion of a furin cleavage site at the S1/S2 boundary in the SARS-CoV spike protein utilized an optimal ‘RRSRR’ sequence in pseudotype systems (Belouzard et al., 2009; Follis et al., 2006). Further, there is no evidence of prior research at the WIV involving the artificial insertion of complete furin cleavage sites into coronaviruses.

The recurring P681H/R substitution in the proline (P) residue preceding the SARS-CoV-2 furin cleavage site improves cleavage of the spike protein and is another signature of ongoing human adaptation of the virus (Peacock et al., 2021a). The SARS-CoV-2 furin site is also lost under standard cell culture conditions involving Vero E6 cells (Ogando et al., 2020; Peacock et al., 2021b), as is true of HCoV-OC43 (Follis et al., 2006). The presence of two adjacent CGG codons for arginine in the SARS-CoV-2 furin cleavage site is similarly not indicative of genetic engineering (Maxmen and Mallapaty, 2021). Although the CGG codon is rare in coronaviruses, it is observed in SARS-CoV, SARS-CoV-2 and other human coronaviruses at comparable frequencies (Maxmen and Mallapaty, 2021). Further, if low-fitness codons had been artificially inserted into the virus genome they would have been quickly selected against during SARS-CoV-2 evolution, yet both CGG codons are more than 99.8% conserved among the >2,300,000 near-complete SARS-CoV-2 genomes sequenced to date, indicative of strong functional constraints (**Supplementary Information, Table S1**).

Conclusions

As for the vast majority of human viruses, the most parsimonious explanation for the origin of SARS-CoV-2 is a zoonotic event. The documented epidemiological history of the virus is comparable to previous animal market-associated outbreaks of coronaviruses with a simple route for human exposure. The contact tracing of SARS-CoV-2 to markets in Wuhan exhibits striking similarities to the early spread of SARS-CoV to markets in Guangdong, where humans infected early in the epidemic lived near or worked in animal markets. Zoonotic spillover by definition selects for viruses able to infect humans. Although strong safeguards should be consistently employed to minimize the likelihood of laboratory accidents in virological research, those laboratory escapes documented to date have almost exclusively involved viruses brought into laboratories specifically because of their known human infectivity.

There is currently no evidence that SARS-CoV-2 has a laboratory origin. There is no evidence that any early cases had any connection to the WIV, in contrast to the clear epidemiological links to animal markets in Wuhan, nor evidence that the WIV possessed or worked on a progenitor of SARS-CoV-2 prior to the pandemic. The suspicion that SARS-CoV-2 might have a laboratory origin stems from the coincidence that it was first detected in a city that houses a major virological laboratory that studies coronaviruses. Wuhan is the largest city in central China with multiple animal markets and is a major hub for travel and commerce, well connected to other areas both within China and internationally. The link to Wuhan therefore more likely reflects the fact that pathogens often require heavily populated areas to become established (Pekar et al., 2021).

We contend that although the animal reservoir for SARS-CoV-2 has not been identified and the key species may not have been tested, in contrast to other scenarios there is substantial body of scientific evidence supporting a zoonotic origin. While the possibility of a laboratory accident cannot be entirely dismissed, and may be near impossible to falsify, this conduit for emergence is highly unlikely relative to the numerous and repeated human-animal contacts that occur routinely in the wildlife trade. Failure to comprehensively investigate the zoonotic origin through collaborative and carefully coordinated studies would leave the world vulnerable to future pandemics arising from the same human activities that have repeatedly put us on a collision course with novel viruses.

Acknowledgements

ECH is supported by an Australian Research Council Australian Laureate Fellowship (FL170100022). SAG is supported by the National Institutes of Health F32AI152341. JOW acknowledges support from the National Institutes of Health (AI135992). ALR acknowledges that VIDO receives operational funding from the Canada Foundation for Innovation - Major Science Initiatives Fund and from the Government of Saskatchewan through Innovation Saskatchewan and the Ministry of Agriculture. DLR acknowledges support of the Medical Research Council (MC_UU_12014/12) and the Wellcome Trust (220977/Z/20/Z). SJA acknowledges funding from the National Institute of Allergy and Infectious Diseases (R01AI149693). WB receives support from the Wellcome Trust (Z/205100 and Z/200187), BBSRC (BB/S008292) and MRC (MR/W005611/1). MFB acknowledges funding from the Bill and Melinda Gates Foundation (INV-005517). JLG is supported by a New Zealand Royal Society Rutherford Discovery Fellowship (RDF-20-UOO-007). JLL is supported by the National Institutes of Health (R01AI141607, R21AI139738) and the National Science Foundation (Grant no. 2029949). SJN is supported by a Wellcome Trust Senior Fellowship (WT098049AIA), the Medical Research Council, and the Huo Family Charitable Foundation 2. TS was funded by the Austrian Science Fund (FWF) grant number P 28183. SRW is supported by the National Institutes of Health (R01AI140442, R01AI104887, R21AI138564, R21AI157147), as well as the Penn Center for Research on Coronaviruses and Other Emerging Pathogens. MW is supported by Bill and Melinda Gates Foundation INV004212 and the Arizona Board of Regents. KGA acknowledges support from the National Institutes of Health (U19AI135995, U01AI151812, and UL1TR002550). RFG is supported by the National Institutes of Health (R01AI132223, R01AI132244, U19AI135995, U54HG007480, U19AI142790, U01AI151812), the Coalition for Epidemic Preparedness Innovations (INTU1901 and ESEP1904) and the European & Developing Countries Clinical Trials Partnership (RIA2019LV-3053). AR acknowledges the support of the Wellcome Trust (Collaborators Award 206298/Z/17/Z – ARTIC network) and the European Research Council (grant agreement no. 725422 – ReservoirDOCS). We gratefully acknowledge the authors and the laboratories responsible for the genome sequence data shared

via the GISAID Initiative, and provide a complete acknowledgement table for the data used here in the **supplementary information**.

Declaration of Interests

ECH is an Honorary Visiting Professor at Fudan University (Shanghai Public Health Clinical Center), Shanghai, China, and between 2014-2020 was a Guest Professor at the Chinese Center for Disease Control and Prevention, Beijing, China. These affiliations are only used in papers co-authored with Prof. Yong-Zhen Zhang (Shanghai Public Health Clinical Center) and involve no formal appointment, no duties and no remuneration nor research funding. JOW receives funding from the U.S. Centers for Disease Control and Prevention (ongoing) via grants and contracts to his institution unrelated to this research. SRW consults for Immunome and Ocugen. AR, ALR, MFB, SAG, and KGA have received consulting fees and compensated expert testimony on SARS-CoV-2 and the COVID-19 pandemic. RFG is co-founder of Zalgen Labs.

References

- Al, H.E.Y. et al. (2004). Surveillance of SARS coronavirus among wild animal sold in Dongmen market in Shenzhen city. *Jbjc* 19, 287–291.
- Andersen, K.G., Rambaut, A., Lipkin, W.I., Holmes, E.C., and Garry, R.F. (2020). The proximal origin of SARS-CoV-2. *Nat. Med.* 26, 450–452.
- Bao, L., Deng, W., Huang, B., Gao, H., Liu, J., Ren, L., Wei, Q., Yu, P., Xu, Y., Qi, F., et al. (2020). The pathogenicity of SARS-CoV-2 in hACE2 transgenic mice. *Nature* 583, 830–833.
- Belouzard, S., Chu, V.C., and Whittaker, G.R. (2009). Activation of the SARS coronavirus spike protein via sequential proteolytic cleavage at two distinct sites. *Proc. Natl. Acad. Sci. USA.* 106, 5871–5876.
- Blow, J.A., Dohm, D.J., Negley, D.L., and Mores, C.N. (2004). Virus inactivation by nucleic acid extraction reagents. *J. Virol. Meth.* 119, 195–198.
- Boni, M.F., Lemey, P., Jiang, X., Lam, T.T.-Y., Perry, B.W., Castoe, T.A., Rambaut, A., and

- Robertson, D.L. (2020). Evolutionary origins of the SARS-CoV-2 sarbecovirus lineage responsible for the COVID-19 pandemic. *Nat. Microbiol.* 5, 1408–1417.
- Cai, Y., Zhang, J., Xiao, T., Lavine, C.L., Rawson, S., Peng, H., Zhu, H., Anand, K., Tong, P., Gautam, A., et al. (2021). Structural basis for enhanced infectivity and immune evasion of SARS-CoV-2 variants. *Science*. doi:10.1126/science.abi9745.
- Centers for Disease Control and Prevention (CDC) (2003). Prevalence of IgG antibody to SARS-associated coronavirus in animal traders - Guangdong Province, China, 2003. *MMWR Morb. Mortal. Wkly. Rep.* 52, 986–987.
- Cohen, J. (2020). Wuhan coronavirus hunter Shi Zhengli speaks out. *Science* 369, 487-488.
- Conceicao, C., Thakur, N., Human, S., Kelly, J.T., Logan, L., Bialy, D., Bhat, S., Stevenson-Leggett, P., Zagrajek, A.K., Hollinghurst, P., et al. (2020). The SARS-CoV-2 Spike protein has a broad tropism for mammalian ACE2 proteins. *PLoS Biol.* 18, e3001016.
- Davidson, A.D., Williamson, M.K., Lewis, S., Shoemark, D., Carroll, M.W., Heesom, K.J., Zambon, M., Ellis, J., Lewis, P.A., Hiscox, J.A., et al. (2020). Characterisation of the transcriptome and proteome of SARS-CoV-2 reveals a cell passage induced in-frame deletion of the furin-like cleavage site from the spike glycoprotein. *Genome Med.* 12, 68.
- Deng, S., Xing, K., and He, X. (2021). Mutation signatures inform the natural host of SARS-CoV-2. *bioRxiv*. doi:10.1101/2021.07.05.451089.
- Dicken, S.J., Murray, M.J., Thorne, L.G., Reuschl, A.-K., Forrest, C., Ganeshalingham, M., Muir, L., Kalemera, M.D., Palor, M., McCoy, L.E., et al. (2021). Characterisation of B.1.1.7 and Pangolin coronavirus spike provides insights on the evolutionary trajectory of SARS-CoV-2. *bioRxiv*. doi:10.1101/2021.03.22.436468.
- Dinnon, K.H., 3rd, Leist, S.R., Schäfer, A., Edwards, C.E., Martinez, D.R., Montgomery, S.A., West, A., Yount, B.L., Jr, Hou, Y.J., Adams, L.E., et al. (2020). A mouse-adapted model of SARS-CoV-2 to test COVID-19 countermeasures. *Nature* 586, 560–566.
- Follis, K.E., York, J., and Nunberg, J.H. (2006). Furin cleavage of the SARS coronavirus spike glycoprotein enhances cell-cell fusion but does not affect virion entry. *Virology* 350, 358–369.

- Freuling, C.M., Breithaupt, A., Müller, T., Sehl, J., Balkema-Buschmann, A., Rissmann, M., Klein, A., Wylezich, C., Höper, D., Wernike, K., et al. (2020). Susceptibility of raccoon dogs for experimental SARS-CoV-2 infection. *Emerg. Infect. Dis.* 26, 2982–2985.
- Gallaher, W.R. (2020). A palindromic RNA sequence as a common breakpoint contributor to copy-choice recombination in SARS-COV-2. *Arch. Virol.* 165, 2341–2348.
- Ge, X., Li, Y., Yang, X., Zhang, H., Zhou, P., Zhang, Y., and Shi, Z. (2012). Metagenomic analysis of viruses from bat fecal samples reveals many novel viruses in insectivorous bats in China. *J. Virol.* 86, 4620–4630.
- Ge, X.-Y., Li, J.-L., Yang, X.-L., Chmura, A.A., Zhu, G., Epstein, J.H., Mazet, J.K., Hu, B., Zhang, W., Peng, C., et al. (2013). Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor. *Nature* 503, 535–538.
- Geddes, A.M. (2006). The history of smallpox. *Clin. Dermatol.* 24, 152–157.
- Gombold, J.L., Hingley, S.T., and Weiss, S.R. (1993). Fusion-defective mutants of mouse hepatitis virus A59 contain a mutation in the spike protein cleavage signal. *J. Virol.* 67, 4504–4512.
- Gu, H., Chen, Q., Yang, G., He, L., Fan, H., Deng, Y.-Q., Wang, Y., Teng, Y., Zhao, Z., Cui, Y., et al. (2020). Adaptation of SARS-CoV-2 in BALB/c mice for testing vaccine efficacy. *Science* 369, 1603–1607.
- Guan, Y., Zheng, B.J., He, Y.Q., Liu, X.L., Zhuang, Z.X., Cheung, C.L., Luo, S.W., Li, P.H., Zhang, L.J., Guan, Y.J., et al. (2003). Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. *Science* 302, 276–278.
- de Haan, C.A.M., Haijema, B.J., Schellen, P., Wichgers Schreur, P., te Lintelo, E., Vennema, H., and Rottier, P.J.M. (2008). Cleavage of group 1 coronavirus spike proteins: how furin cleavage is traded off against heparan sulfate binding upon cell culture adaptation. *J. Virol.* 82, 6078–6083.
- Hassan, A.O., Case, J.B., Winkler, E.S., Thackray, L.B., Kafai, N.M., Bailey, A.L., McCune, B.T., Fox, J.M., Chen, R.E., Alsoussi, W.B., et al. (2020). A SARS-CoV-2 infection model in

mice demonstrates protection by neutralizing antibodies. *Cell* 182, 744–753.e4.

Hu, B., Zeng, L.-P., Yang, X.-L., Ge, X.-Y., Zhang, W., Li, B., Xie, J.-Z., Shen, X.-R., Zhang, Y.-Z., Wang, N., et al. (2017). Discovery of a rich gene pool of bat SARS-related coronaviruses provides new insights into the origin of SARS coronavirus. *PLoS Pathog.* 13, e1006698.

Israelow, B., Song, E., Mao, T., Lu, P., Meir, A., Liu, F., Alfajaro, M.M., Wei, J., Dong, H., Homer, R.J., et al. (2020). Mouse model of SARS-CoV-2 reveals inflammatory role of type I interferon signaling. *J. Exp. Med.* 217, e20201241.

Khan, A., Zia, T., Suleman, M., Khan, T., Ali, S.S., Abbasi, A.A., Mohammad, A., and Wei, D.-Q. (2021). Higher infectivity of the SARS-CoV-2 new variants is associated with K417N/T, E484K, and N501Y mutants: An insight from structural data. *J. Cell. Physiol.* doi:10.1002/jcp.30367.

Kirchdoerfer, R.N., Cottrell, C.A., Wang, N., Pallesen, J., Yassine, H.M., Turner, H.L., Corbett, K.S., Graham, B.S., McLellan, J.S., and Ward, A.B. (2016). Pre-fusion structure of a human coronavirus spike protein. *Nature* 531, 118–121.

Klimstra, W.B., Tilston-Lunel, N.L., Nambulli, S., Boslett, J., McMillen, C.M., Gilliland, T., Dunn, M.D., Sun, C., Wheeler, S.E., Wells, A., et al. (2020). SARS-CoV-2 growth, furin-cleavage-site adaptation and neutralization using serum from acutely infected hospitalized COVID-19 patients. *J. Gen. Virol.* 101, 1156–1169.

Korber, B., Fischer, W.M., Gnanakaran, S., Yoon, H., Theiler, J., Abfalterer, W., Hengartner, N., Giorgi, E.E., Bhattacharya, T., Foley, B., et al. (2020). Tracking changes in SARS-CoV-2 spike: evidence that D614G increases infectivity of the COVID-19 virus. *Cell* 182, 812–827.e19.

Kuzmina, A., Khalaila, Y., Voloshin, O., Keren-Naus, A., Boehm-Cohen, L., Raviv, Y., Shemer-Avni, Y., Rosenberg, E., and Taube, R. (2021). SARS-CoV-2 spike variants exhibit differential infectivity and neutralization resistance to convalescent or post-vaccination sera. *Cell Host Microbe* 29, 522–528.e2.

Latinne, A., Hu, B., Olival, K.J., Zhang, L., Li, H., Chmura, A. A., Field, H. E., Zambrana-Torrel, C., Epstein, J. H., Li, B., et al. (2020). Origin and cross-species transmission of bat

coronaviruses in China. *Nat. Commun.* *11*, 4235.

Lednický, J.A., Tagliamonte, M.S., White, S.K., Elbadry, M.A., Alam, M.M., Stephenson, C.J., Bonny, T.S., Loeb, J.C., Telisma, T., Chavannes, S., et al. (2021). Emergence of porcine delta-coronavirus pathogenic infections among children in Haiti through independent zoonoses and convergent evolution. *medRxiv*. doi:10.1101/2021.03.19.21253391.

Leist, S.R., Dinno, K.H., 3rd, Schäfer, A., Tse, L.V., Okuda, K., Hou, Y.J., West, A., Edwards, C.E., Sanders, W., Fritch, E.J., et al. (2020). A mouse-adapted SARS-CoV-2 induces acute lung injury and mortality in standard laboratory mice. *Cell* *183*, 1070–1085.e12.

Li, L.-L., Wang, J.-L., Ma, X.-H., Li, J.-S., Yang, X.-F., Shi, W.-F., and Duan, Z.-J. (2021). A novel SARS-CoV-2 related virus with complex recombination isolated from bats in Yunnan province, China. *bioRxiv*. doi:10.1101/2021.03.17.435823.

Lim, P.L., Kurup, A., Gopalakrishna, G., Chan, K.P., Wong, C.W., Ng, L.C., Se-Thoe, S.Y., Oon, L., Bai, X., Stanton, L.W., et al. (2004). Laboratory-acquired severe acute respiratory syndrome. *N. Engl. J. Med.* *350*, 1740–1745.

Liu, M., Deng, L., Wang, D., and Jiang, T. (2020a). Influenza activity during the outbreak of coronavirus disease 2019 in Chinese mainland. *Biosaf Health* *2*, 206–209.

Liu, Y., Liu, J., Plante, K.S., Plante, J.A., Xie, X., Zhang, X., Ku, Z., An, Z., Scharon, D., Schindewolf, C., et al. (2021). The N501Y spike substitution enhances SARS-CoV-2 transmission. *bioRxiv*. doi: 10.1101/2021.03.08.434499.

Liu, Z., Zheng, H., Lin, H., Li, M., Yuan, R., Peng, J., Xiong, Q., Sun, J., Li, B., Wu, J., et al. (2020b). Identification of common deletions in the spike protein of Severe Acute Respiratory Syndrome coronavirus 2. *J. Virol.* *94*, e00790-20.

Lytras, S., Hughes, J., Martin, D., de Klerk, A., Lourens, R., Kosakovsky Pond, S.L., Xia, W., Jiang, X., and Robertson, D.L. Exploring the natural origins of SARS-CoV-2 in the light of recombination. *bioRxiv*. doi:10.1101/2021.01.22.427830.

Maxmen, A., and Mallapaty, S. (2021). The COVID lab-leak hypothesis: what scientists do and don't know. *Nature* *594*, 313–315.

- Menachery, V.D., Yount, B.L., Jr, Debbink, K., Agnihothram, S., Gralinski, L.E., Plante, J.A., Graham, R.L., Scobey, T., Ge, X.-Y., Donaldson, E.F., et al. (2015). A SARS-like cluster of circulating bat coronaviruses shows potential for human emergence. *Nat. Med.* *21*, 1508–1513.
- Ogando, N.S., Dalebout, T.J., Zevenhoven-Dobbe, J.C., Limpens, R.W.A.L., van der Meer, Y., Caly, L., Druce, J., de Vries, J.J.C., Kikkert, M., Bárcena, M., et al. (2020). SARS-coronavirus-2 replication in Vero E6 cells: replication kinetics, rapid adaptation and cytopathology. *J. Gen. Virol.* *101*, 925–940.
- Otto, S.P., Day, T., Arino, J., Colijn, C., Dushoff, J., Li, M., Mechai, S., Van Domselaar, G., Wu, J., Earn, D.J.D., et al. (2021). The origins and potential future of SARS-CoV-2 variants of concern in the evolving COVID-19 pandemic. *Curr. Biol.* *31*, R918-R929.
- Oude Munnink, B.B., Sikkema, R.S., Nieuwenhuijse, D.F., Molenaar, R.J., Munger, E., Molenkamp, R., van der Spek, A., Tolsma, P., Rietveld, A., Brouwer, M., et al. (2021). Transmission of SARS-CoV-2 on mink farms between humans and mink and back to humans. *Science* *371*, 172–177.
- Parry, J. (2004). Breaches of safety regulations are probable cause of recent SARS outbreak, WHO says. *BMJ* *328*, 1222.
- Peacock, T.P., Sheppard, C.M., Brown, J.C., Goonawardane, N., Zhou, J., Whiteley, M., PHE Virology Consortium, de Silva, T.I., and Barclay, W.S. (2021a). The SARS-CoV-2 variants associated with infections in India, B.1.617, show enhanced spike cleavage by furin. *bioRxiv*. doi:10.1101/2021.05.28.446163.
- Peacock, T.P., Goldhill, D.H., Zhou, J., Baillon, L., Frise, R., Swann, O.C., Kugathasan, R., Penn, R., Brown, J.C., Sanchez-David, R.Y., et al. (2021b). The furin cleavage site in the SARS-CoV-2 spike protein is required for transmission in ferrets. *Nat Microbiol.* doi:10.1038/s41564-021-00908-w.
- Pekar, J., Worobey, M., Moshiri, N., Scheffler, K., and Wertheim, J.O. (2021). Timing the SARS-CoV-2 index case in Hubei province. *Science* *372*, 412–417.
- Piplani, S., Singh, P.K., Winkler, D.A., and Petrovsky, N. (2021). *In silico* comparison of SARS-

CoV-2 spike protein-ACE2 binding affinities across species and implications for virus origin. *Sci. Rep.* *11*, 13063.

Rambaut, A., Holmes, E.C., O'Toole, Á., Hill, V., McCrone, J.T., Ruis, C., du Plessis, L., and Pybus, O.G. (2020). A dynamic nomenclature proposal for SARS-CoV-2 lineages to assist genomic epidemiology. *Nat Microbiol* *5*, 1403–1407.

Rathnasinghe, R., Strohmeier, S., Amanat, F., Gillespie, V.L., Krammer, F., García-Sastre, A., Coughlan, L., Schotsaert, M., and Uccellini, M.B. (2020). Comparison of transgenic and adenovirus hACE2 mouse models for SARS-CoV-2 infection. *Emerg. Microbes Infect.* *9*, 2433–2445.

Ristanović, E.S., Kokoškov, N.S., Crozier, I., Kuhn, J.H., and Gligić, A.S. (2020). A forgotten episode of Marburg virus disease: Belgrade, Yugoslavia, 1967. *Microbiol. Mol. Biol. Rev.* *84*, e00095–19.

Rozo, M., and Gronvall, G.K. (2015). The reemergent 1977 H1N1 strain and the gain-of-function debate. *mBio* *6*, e01013–e01015.

Sasaki, M., Uemura, K., Sato, A., Toba, S., Sanaki, T., Maenaka, K., Hall, W.W., Orba, Y., and Sawa, H. (2021). SARS-CoV-2 variants with mutations at the S1/S2 cleavage site are generated in vitro during propagation in TMPRSS2-deficient cells. *PLoS Pathog.* *17*, e1009233.

Senio, K. (2003). Recent Singapore SARS case a laboratory accident. *Lancet Infect. Dis.* *3*, 679.

Simmonds, P. (2020). Rampant C→U hypermutation in the genomes of SARS-CoV-2 and other coronaviruses: causes and consequences for their short- and long-term evolutionary trajectories. *mSphere* *5*, e00408–e00420.

Starr, T.N., Greaney, A.J., Hilton, S.K., Ellis, D., Crawford, K.H.D., Dingens, A.S., Navarro, M.J., Bowen, J.E., Tortorici, M.A., Walls, A.C., et al. (2020). Deep mutational scanning of SARS-CoV-2 receptor binding domain reveals constraints on folding and ACE2 binding. *Cell* *182*, 1295–1310.e20.

Sun, S., Gu, H., Cao, L., Chen, Q., Yang, G., Li, R.-T., Fan, H., Ye, Q., Deng, Y.-Q., Song, X., et al. (2020a). Characterization and structural basis of a lethal mouse-adapted SARS-CoV-2.

bioRxiv. doi:10.1101/2020.11.10.377333.

Sun, S.-H., Chen, Q., Gu, H.-J., Yang, G., Wang, Y.-X., Huang, X.-Y., Liu, S.-S., Zhang, N.-N., Li, X.-F., Xiong, R., et al. (2020b). A mouse model of SARS-CoV-2 infection and pathogenesis. *Cell Host Microbe* 28, 124–133.e4.

Tu, C., Crameri, G., Kong, X., Chen, J., Sun, Y., Yu, M., Xiang, H., Xia, X., Liu, S., Ren, T., et al. (2004). Antibodies to SARS coronavirus in civets. *Emerg. Infect. Dis.* 10, 2244–2248.

van Aart, A.E., Velkers, F.C., Fischer, E.A.J., Broens, E.M., Egberink, H., Zhao, S., Engelsma, M., Hakze-van der Honing, R.W., Harders, F., de Rooij, M.M.T., et al. (2021). SARS-CoV-2 infection in cats and dogs in infected mink farms. *Transbound. Emerg. Dis.* doi:10.1111/tbed.14173.

Vlasova, A.N., Diaz, A., Damtie, D., Xiu, L., Toh, T.-H., Lee, J.S.-Y., Saif, L.J., and Gray, G.C. (2021). Novel canine coronavirus isolated from a hospitalized patient with pneumonia in east Malaysia. *Clin. Infect. Dis.* doi:10.1093/cid/ciab456.

Volz, E., Hill, V., McCrone, J.T., Price, A., Jorgensen, D., O'Toole, Á., Southgate, J., Johnson, R., Jackson, B., Nascimento, F.F., et al. (2021). Evaluating the effects of SARS-CoV-2 spike mutation D614G on transmissibility and pathogenicity. *Cell* 184, 64–75.e11.

Wacharapluesadee, S., Tan, C.W., Maneerorn, P., Duengkae, P., Zhu, F., Joyjinda, Y., Kaewpom, T., Chia, W.N., Ampoot, W., Lim, B.L., et al. (2021). Evidence for SARS-CoV-2 related coronaviruses circulating in bats and pangolins in Southeast Asia. *Nat. Commun.* 12, 1–9.

Wan, Y., Shang, J., Graham, R., Baric, R.S., and Li, F. (2020). Receptor recognition by the novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS coronavirus. *J. Virol.* 94, e00127–20.

Wang M., Jing H.-Q., Xu H.-F., Jiang X.-G., Kan B., Liu Q.-Y., Wan K.-L., Cui B.-Y., Zheng H., Cui Z.-G., et al. (2005). Surveillance on severe acute respiratory syndrome associated coronavirus in animals at a live animal market of Guangzhou in 2004. *Zhonghua Liu Xing Bing Xue Za Zhi* 26, 84–87.

Wang, N., Li, S.-Y., Yang, X.-L., Huang, H.-M., Zhang, Y.-J., Guo, H., Luo, C.-M., Miller, M.,

Zhu, G., Chmura, A.A., et al. (2018). Serological evidence of bat SARS-related coronavirus infection in humans, China. *Virol. Sin.* 33, 104–107.

WHO (2021). WHO-convened global study of origins of SARS-CoV-2: China Part (World Health Organization).

Wong, Y.C., Lau, S.Y., To, K.K.W., Mok, B.W.Y., Li, X., Wang, P., Deng, S., Woo, K.F., Du, Z., Li, C., et al. (2020). Natural transmission of bat-like Severe Acute Respiratory Syndrome Coronavirus 2 without proline-arginine-arginine-alanine variants in Coronavirus Disease 2019 patients. *Clin. Infect. Dis.* 73, e437-e444.

Woo, P.C.Y., Lau, S.K.P., Chu, C.-M., Chan, K.-H., Tsoi, H.-W., Huang, Y., Wong, B.H.L., Poon, R.W.S., Cai, J.J., Luk, W.-K., et al. (2005). Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. *J. Virol.* 79, 884–895.

Xiao, K., Zhai, J., Feng, Y., Zhou, N., Zhang, X., Zou, J.-J., Li, N., Guo, Y., Li, X., Shen, X., et al. (2020). Isolation of SARS-CoV-2-related coronavirus from Malayan pangolins. *Nature* 583, 286–289.

Xiao, X., Newman, C., Buesching, C.D., Macdonald, D.W., and Zhou, Z.-M. (2021). Animal sales from Wuhan wet markets immediately prior to the COVID-19 pandemic. *Sci. Rep.* 11, 1–7.

Xu, R.-H., He, J.-F., Evans, M.R., Peng, G.-W., Field, H.E., Yu, D.-W., Lee, C.-K., Luo, H.-M., Lin, W.-S., Lin, P., et al. (2004). Epidemiologic clues to SARS origin in China. *Emerg. Infect. Dis.* 10, 1030–1037.

Yang, X.-L., Hu, B., Wang, B., Wang, M.-N., Zhang, Q., Zhang, W., Wu, L.-J., Ge, X.-Y., Zhang, Y.-Z., Daszak, P., et al. (2015). Isolation and characterization of a novel bat coronavirus closely related to the direct progenitor of Severe Acute Respiratory Syndrome coronavirus. *J. Virol.* 90, 3253–3256.

Zhan, S.H., Deverman, B.E., and Chan, Y.A. (2020). SARS-CoV-2 is well adapted for humans. What does this mean for re-emergence? *bioRxiv* doi:2020.05.01.073262.

Zhou, H., Ji, J., Chen, X., Bi, Y., Li, J., Wang, Q., Hu, T., Song, H., Zhao, R., Chen, Y., et al.

(2021). Identification of novel bat coronaviruses sheds light on the evolutionary origins of SARS-CoV-2 and related viruses. *Cell*. doi:10.1016/j.cell.2021.06.008.

Zhou, H., Chen, X., Hu, T., Li, J., Song, H., Liu, Y., Liu, D., Yang, J., Holmes, E.C., Hughes, A.C., et al. (2020). A novel bat coronavirus closely related to SARS-CoV-2 contains natural insertions at the S1/S2 cleavage site of the spike protein. *Curr. Biol.* *30*, 1-8.

Zhou, P., Yang, X.L., Wang, X.G., Hu, B., Zhang, L., Zhang, W., Si, H.-R., Zhu, Y., Li, B., Huang, C.-L., et al. (2020). A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* *579*, 270–273.

Zhu, X., Mannar, D., Srivastava, S.S., Berezuk, A.M., Demers, J.-P., Saville, J.W., Leopold, K., Li, W., Dimitrov, D.S., Tuttle, K.S., et al. (2021a). Cryo-electron microscopy structures of the N501Y SARS-CoV-2 spike protein in complex with ACE2 and 2 potent neutralizing antibodies. *PLoS Biol.* *19*, e3001237.

Zhu, Y., Feng, F., Hu, G., Wang, Y., Yu, Y., Zhu, Y., Xu, W., Cai, X., Sun, Z., Han, W., et al. (2021b). A genome-wide CRISPR screen identifies host factors that regulate SARS-CoV-2 entry. *Nat. Commun.* *12*, 961.

Figure Legends

Figure 1. Phylogenetic and epidemiological data on the early COVID-19 pandemic in Wuhan. (A) Phylogenetic tree of early SARS-CoV-2 genomes sampled from Wuhan during December 2019-January 2020. The split between lineages A and B is labelled with the coordinates and base of the two differentiating nucleotide mutations. Cases with a known association to the Huanan or other markets are denoted by symbols (reported in WHO, 2021). (B) Map of districts of Wuhan showing the location of markets, the Wuhan National Biosafety Laboratory at the Zhengdian Scientific Park of the Wuhan Institute of Virology (denoted WIV), where the coronavirus isolation and culture work of Dr. Shi Zhengli is performed, and the earliest known cases. (C-E) Location of recorded COVID-19 cases in Wuhan from 8th December to 31st December 2019. Cases with a home address outside of Wuhan city are not shown. (F-H) Map of districts of Wuhan indicating the first record of excess deaths due to pneumonia (shaded green) from 15th January 2020. Case and excess death data were extracted and redrawn from figures provided in WHO, 2021. For more details see **Supplementary Information**.

Figure 2. Evolution of the furin cleavage site (FCS) in the spike protein of betacoronaviruses. (A) Sequence alignment of the region around the FCS in SARS-CoV-2 (NCBI accession MN908947) and bat coronavirus RaTG13 (NCBI accession MN996532) showing that the former was the result of an out-of-frame nucleotide sequence insertion. (B) Amino acid sequence alignment of the FCS region in representative members of the different subgenera of betacoronaviruses, highlighting the evolutionary volatility of this site and that the relevant amino acid motif (RRAR) in SARS-CoV-2 is functionally suboptimal. The residues predicted to be O-linked glycans are also marked. For more details see **Supplementary Information**.

The Origins of SARS-CoV-2: A Critical Review

Supplementary Information

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Figure 1 Phylogenetic tree

SARS-CoV-2 genome sequences were downloaded from the GISAID EpiCoV database (<http://gisaid.org>). All complete and high coverage genomes from Wuhan, China with collection dates from December 2019 to January 2020 were downloaded. Genomes were pairwise aligned to the reference genome, ‘Wuhan/Hu-1/2019’ (NCBI accession MN908947) using Minimap2 [1] and the 5’ and 3’ untranslated regions were masked to avoid areas of low sequencing coverage. A maximum likelihood phylogenetic tree was estimated using IQ-TREE2 [2] under the Jukes-Cantor model of nucleotide substitution. The tree was rooted at the midpoint between lineage A and lineage B.

Three genomes from late January 2020 were removed (‘Wuhan/0126-C94/2020’, ‘Wuhan/0126-C100/2020’, ‘Wuhan/0126-C93/2020’ – GISAID accessions EPI_ISL_493180, EPI_ISL_493182, EPI_ISL_493179, respectively) because although they had the mutation 8782T indicative of lineage A, they did not have the corresponding 28144C mutation. One of these sequences, ‘Wuhan/0126-C93/2020’, shares a mutation (13402G) with a lineage A genome from the same collection date and laboratory (‘Wuhan/0126-C77/2020’). It is likely that the nucleotide at 28144 has been called as the reference allele (28144T - using the Wuhan-Hu-1 reference genome).

Information about 13 early cases linked to genomes was collected from published work and Tables 6 and 7 from the WHO report [3]. Where there were discrepancies, the published reports were given priority. In particular, the case in Tables 6 and 7 with the earliest onset date (2019-12-08) seems to have been mistakenly linked to a genome (see Table S2, note 1). Where multiple genomes were linked to the same case in Table 6 of ref. 3, only one representative was included (Table S2).

Figure 1 Map Locations

Xiao, X., Newman, C., Buesching, C.D., Macdonald, D.W., and Zhou, Z.-M. (2021). Animal sales from Wuhan wet markets immediately prior to the COVID-19 pandemic. *Sci. Rep.* 11, 1–7. [4]

- Baishazhou market, Li Shui Lu, Hongshan Qu, Wuhan Shi, Hubei Sheng, China: 30.4626°N 114.2565°E
- Qiyimen Shengxian farmer’s market, 588 Zhongshan Rd, Wuchang District, Wuhan, Hubei, China: 30.5232°N 114.3096°E
- Huanan Seafood Wholesale Market, Fazhan Ave, Jiangnan District, Wuhan, Hubei, China: 30.6196°N 114.2576°E

“These shops selling live, often wild, animals included two at the Baishazhou market (a large market comprising c. 400 other types of shop), seven at Huanan seafood market (c. 120 other shops), four at Dijiao outdoor pet market (c. 100 other shops), and four at Qiyimen live animal market (c. 40 other shops).”

The Wuhan Institute of Virology (WIV):

- The Wuhan National Biosafety Laboratory at the Zhengdian Scientific Park of the Wuhan Institute of Virology: 30.376389°N, 114.262500°E

Panels b-d: Map data was manually extracted from Fig 17 (Page 157) of the Annexes of ref. 3 using Adobe Illustrator. Because of multiple overlapping points there will be errors in the extraction process. Peripheral districts are: DXH: Dongxihu, CD: Caidian, JX: Jiangxia, HP: Huangpi, XZ: Xinzhou and HN: Hannan.

Panels e-f. Excess mortality from pneumonia by district/governmental areas from Fig. 21 (p. 40) of ref. 3 is indicated for selected dates.

Map data and polygons from OpenStreetMap (<http://openstreetmap.org>) and copyright © OpenStreetMap contributors – see <https://www.openstreetmap.org/copyright> for details.

Figure 2 - Methods.

Panel a: Alignment of the nucleotide sequences encoding the S1/S2 cleavage sites of the spike proteins of SARS-CoV-2 (YP_009724390 and bat Coronavirus RaTG13 (QHR63300.2). The reading frame for the amino acids can be inferred from the variation in the third base of several codons (yellow). Two possible insertions are indicated by capital letters, both of which are out-of-frame (-1 or -2). Numbers represent amino acids of the Spike proteins and nucleotides of the entire genomes.

Panel b: Amino acid alignment of the S1/S2 cleavage sites of selected beta spike proteins. Accession numbers: SARS-CoV-2 YP_009724390, SARS-CoV AAP13441.1, RaTG13 QHR63300.2, RmYN02 EPI_ISL_412977, MERS-CoV AGG22542.1, HKU4 MH002339.1 HKU5 AGP04943.1, HKU5 AGP04943.1, HKU1a ABD75561_1, HKU1b ABD96196_1, OC43 AIX10760.1, Bovine CoV CCE89341.1, HKU24 YP_009113025.1, Chinese *Hipposideros pratti* Bat-betacoronavirus/Zhejiang2013 (HpZJ13) and Nigerian *Hipposideros commersoni* Zaria bat coronavirus (HcNG08). To facilitate the identification of insertions we aligned a conserved cysteine residue (green) and included spikes from viruses that appear to be ancestral to the subgenuses where known. O-linked glycosylation sites were predicted by Net-O-Glyc v. 4.0.

Data files, map data and other supplementary materials are available from <http://github.com/sars-cov-2-origins/critical-review/>

Supplementary Table S1. Codons in the spike furin cleavage site of SARS-CoV-2.

Codon	Amino acid	Residue 682		Residue 683	
		count	proportion	count	proportion
CGG	R	2317072	0.99851	2308542	0.99484
CGT	R	2894	0.00125	3181	0.00137
CGC	R	215	9.26516E-05	52	2.24088E-05
CGA	R	200	8.61876E-05	449	0.000193491
TGG	W	50	2.15469E-05	218	9.39445E-05
AGG	R	34	1.46519E-05	402	0.000173237
CAG	Q	26	1.12044E-05	239	0.000102994
CTG	L	26	1.12044E-05	156	6.72263E-05
CCG	P	2	8.61876E-07	11	4.74032E-06
CAT	H	1	4.30938E-07	1	4.30938E-07
GGG	G	0	0	1	4.30938E-07
AAG	K	0	0	1	4.30938E-07
		2320520	0.00144*	2313252	0.00177*
			99.86%**		99.82%**

* Proportion of non-CGG arginine codons

** Percentage CGG relative to all arginine codons

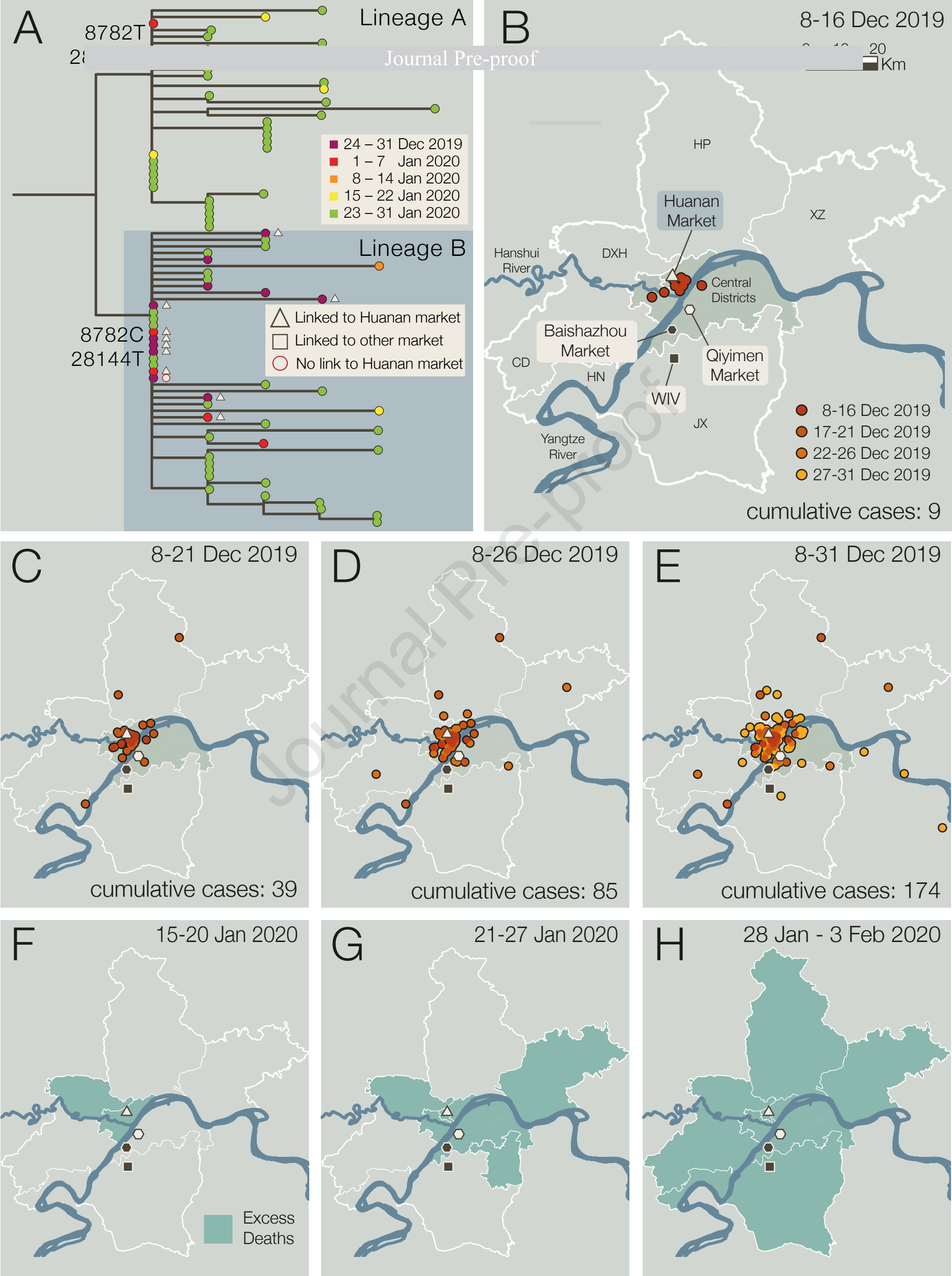
Supplementary Table S2. Early cases linked to genome sequences.

Onset date	Collection date	age/sex	Sequence name	GISAID id	Relation to the Huanan market	reference	
2019-12-16	2019-12-30	41M	Wuhan/IPBCAMS-WH-03/2019	EPI_ISL_403930	none	[5]	Note 1
2019-12-15	2019-12-24	65M	Wuhan/IPBCAMS-WH-01/2019	EPI_ISL_402123	vendor	[5]	
2019-12-17	2019-12-26	44M	Wuhan/WH01/2019	EPI_ISL_406798	purchaser	[3]	
2019-12-19	2019-12-30	32M	Wuhan/HBCDC-HB-02/2019	EPI_ISL_412898	vendor	[3]	Note 2
2019-12-20	2019-12-30	61M	Wuhan/IPBCAMS-WH-05/2020	EPI_ISL_403928	purchaser	[5]	Note 1
2019-12-20	2019-12-26	41M	Wuhan/Hu-1/2019	EPI_ISL_402125	worker	[6]	
2019-12-20	2020-01-02	39M	Wuhan/WHU01/2020	EPI_ISL_406716	vendor	[7]	
2019-12-20	2019-12-30	56M	Wuhan/IME-WH04/2019	EPI_ISL_529216	vendor	[3]	Note 3
					Contact with Huanan Market		
2019-12-22	2020-01-02	21F	Wuhan/WHU02/2020	EPI_ISL_406717	staff	[7]	
2019-12-23	2019-12-30	49F	Wuhan/IPBCAMS-WH-02/2019	EPI_ISL_403931	vendor	[5]	Note 1
2019-12-23	2019-12-30	52F	Wuhan/IPBCAMS-WH-04/2019	EPI_ISL_403929	vendor	[5]	Note 1
2019-12-23	2019-12-30	40M	Wuhan/WIV06/2019	EPI_ISL_402129	vendor	[3]	
					visitor to another		
2019-12-26	2019-12-30		Wuhan/IME-WH01/2019	EPI_ISL_529213	market	[3]	

- Note 1: Patient 1,2,3 & 5 from ref. 4 were matched by age/sex and collection date in GISAID entry. Patient 4 was matched by elimination.
- Note 2: Age/sex taken from EPI_ISL_402127 - WIV02 - Linked in Table 6 of ref. 3 to be the same case.
- Note 3: Age/sex taken from EPI_ISL_402130 - WIV07 - Linked in Table 6 of ref. 3 to be the same case.

References

1. Li, H. (2018). Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34, 3094-3100.
2. Minh, B.Q., Schmidt, H.A., Chernomor, O., Schrempf, D., Woodhams, M.D., von Haeseler A., and Lanfear, R. (2021). IQ-TREE 2: New models and efficient methods for phylogenetic inference in the genomic era. *Mol.Biol.Evol.* 37, 1530–1534.
3. WHO (2021). WHO-convened global study of origins of SARS-CoV-2: China Part (World Health Organization). [cited 28 Jun 2021]. Available at: <https://www.who.int/publications/i/item/who-convened-global-study-of-origins-of-sars-cov-2-china-part>.
4. Xiao, X., Newman, C., Buesching, C.D., Macdonald, D.W., and Zhou, Z.-M. (2021). Animal sales from Wuhan wet markets immediately prior to the COVID-19 pandemic. *Sci. Rep.* 11, 1–7.
5. Ren, L.-L., Wang, Y.-M., Wu, Z.-Q., Xiang, Z.-C., Guo, L., Xu, T., Jiang, Y.-Z., Xiong, Y., Li, Y.-J., Li, X.-W. et al., (2020). Identification of a novel coronavirus causing severe pneumonia in human: a descriptive study. *China Med. J.* 133, 1015-1024.
6. Wu, F., Zhao, S., Yu, B., Chen, Y.-M., Wang, W., Song, Z.-G., Hu, Y., Tao, Z.-W., Tian, J.-H., Pei, Y.-Y., et al. (2020). A new coronavirus associated with human respiratory disease in China. *Nature* 579, 265–269.
7. Chen, L., Liu, W., Zhang, Q., Xu, K., Ye, G., Wu, W., Sun, Z., Liu, F., Wu, K., Zhong, B., et al. (2020). RNA based mNGS approach identifies a novel human coronavirus from two individual pneumonia cases in 2019 Wuhan outbreak. *Emerg Microbes Infect.* 9, 313–319.



-2 reading frame insertion

monobasic cleavage site **R**; predicted O-linked glycan **S/T**